Discovery and Evaluation of *N*-Cyclopropyl-2,4-difluoro-5-((2-(pyridin-2-ylamino)thiazol-5ylmethyl)amino)benzamide (BMS-605541), a Selective and Orally Efficacious Inhibitor of Vascular Endothelial Growth Factor Receptor-2

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Abstract: Substituted 3-((2-(pyridin-2-ylamino)thiazol-5-ylmethyl)amino)benzamides were identified as potent and selective inhibitors of vascular endothelial growth factor receptor-2 (VEGFR-2) kinase activity. The enzyme kinetics associated with the VEGFR-2 inhibition of **14** ($K_i = 49 \pm 9$ nM) confirmed that the aminothiazole-based analogues are competitive with ATP. Analogue **14** demonstrated excellent kinase selectivity, favorable pharmacokinetic properties in multiple species, and robust in vivo efficacy in human lung and colon carcinoma xenograft models.

Tumor-induced angiogenesis, or the formation of new capillary networks between neoplastic cells and endothelial cells of the host,¹ is required for solid tumor growth and metastases.² VEGF,^a a key stimulator of tumor angiogenesis, promotes endothelial cell proliferation, survival, migration, invasion, and differentiation.³ The intrinsic activity responsible for these cellular events is dependent on the highly specific binding of the VEGF ligands to their respective cell surface expressed tyrosine kinase receptors VEGFR-1 (Flt-1), VEGFR-2 (KDR, Flk-1), and VEGFR-3 (Flt-4).⁴ For more than a decade, the VEGF signaling pathway has been the target of extensive drug discovery research aimed at developing therapies that control the abnormal vascular development operative in cancer progression.5 To this end, the chimeric anti-VEGF monoclonal antibody bevacizumab (Genentech) was recently approved to treat patients with metastatic colorectal cancer when combined with standard chemotherapy (IFL/Saltz regimen).⁶ In addition to biologicalbased antiangiogenesis approaches, several small-molecule inhibitors of VEGFR-2 kinase activity have progressed to various stages of preclinical and clinical development.5

We have recently disclosed the structure–activity relationships (SARs) of various ATP-competitive VEGFR-2 inhibitors assembled from the pyrrolo[2,1-*f*][1,2,4]triazine scaffold.⁷ Fourpoint pharmacophore^{8a} and atom pairs^{8b} similarity searches conducted with the potent *N*-methoxybenzamide-substituted pyrrolo[2,1-*f*][1,2,4]triazine inhibitor **1** (VEGFR-2 IC₅₀ = 18 nM)^{7b} identified several *N*-acyl-2-aminothiazole-based cyclindependent kinase (CDK2) inhibitors⁹ from the Bristol-Myers Squibb compound collection. In general, these compounds displayed weak to moderate inhibition of VEGFR-2 kinase activity. For example, N-(5-((5-*tert*-butyloxazol-2-yl)methyl-thio)thiazol-2-yl)-3,4-dimethoxybenzamide (**2**), which demonstrated an IC₅₀ of 50 nM against CDK2, was subsequently found to be a 1900 nM inhibitor of VEGFR-2.



A comparison of the relative binding modes of **1** and **2** in the VEGFR-2 ATP-binding site is provided in Figure 1.^{7b,10} In this model, the pyrrolo[2,1-*f*][1,2,4]triazine and aminothiazole cores are anchored to the hinge region of the adenine binding pocket via a highly conserved hydrogen-bond interaction with the backbone amide-NH of Cys919. The hydroxamate of **1** is believed to be engaged in hydrogen-bond interactions with Asp1046 and Glu885. The overlay of **1** and **2** in this model suggested that attachment of the *N*-methoxybenzamide onto the aminothiazole template may lead to improved potency relative to the initial lead **2**. Herein, we describe the optimization and preclinical assessment of these selective aminothiazole-based VEGFR-2 inhibitors.

To evaluate the aforementioned hypothesis, N-acyl-2aminothiazole and the N-2-pyridinylaminothiazole moiety used to prepare potent pan-Src kinase inhibitors¹¹ and structurally related VEGFR-2 inhibitors¹² were appended via a thiomethyl linker to the N-methoxybenzamide. As depicted in Table 1, the initial N-2-pyridinylaminothiazole analogue 3 displayed potent inhibition of VEGFR-2 kinase activity and VEGF-stimulated human umbilical vein endothelial cell (HUVEC) proliferation. Contrary to the SAR observed in the pyrrolo[2,1-f][1,2,4]triazine series,^{7b} the simple *N*-methylamide analogue **4** retained activity in the biochemical assay. Because of the potential metabolic instability associated with the thiomethyl group, variations in the linker region (X-Y) were pursued. The methylamine analogue 5 was approximately 3-fold less potent than 4 in the VEGFR-2 assay. The ethyl- and *n*-propylbenzamide analogues provided similar biochemical potency relative to 5 (data not shown). Consistent with earlier observations,7b incorporation of a fluorine atom at R¹ of the benzamide ring (i.e., 6) provided a 10-fold increase in intrinsic activity relative to 5. Compound 6 also demonstrated significant antiproliferative activity in the VEGF-driven HUVEC assay. Since the 5-tert-butyloxazolylmethylthio moiety was previously shown to be critical for potent CDK2 inhibition in the aminothiazole series,9 we evaluated and confirmed the selectivity of the current series for VEGFR-2 versus CDK2 (compound 6, $IC_{50} = 5100 \text{ nM}$). Replacement of the methyl group of amide 6 with a cyclopropyl substituent was well tolerated with the amine (Y = NH) and ethyl $(Y = CH_2)$ linkers (i.e., 7 and 8, respectively). Compound 8 was found to be the most potent analogue in the biochemical and cellular

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^{*a*} Abbreviations: VEGF, vascular endothelial growth factor; IFL, irinotecan-5-fluorouracil-leucovorin; HUVEC, human umbilical vein endothelial cell; PK, pharmacokinetics; TFA, trifluoroacetic acid; AUC, area under the curve; V_{ss} , steady-state volume of distribution; Cl, clearance; MRT, mean residence time; TGI, tumor growth inhibition; hERG, human ether-a-gogo-related gene; HEK, human embryonic kidney cells.



Figure 1. Proposed binding model of 1 and 2 in the ATP-binding site of VEGFR-2 kinase.

Table 1. Enzymatic and Cellular Activities^a of N-2-Pyridinylaminothiazole Analogues

| | $R^1 \sim R^2$ |
|-----------------------------------|--------------------|
| NNSY | |
| | `Y``R ³ |
| N[−] | Ö |

| | | | | | | IC ₅₀ , nM | |
|-------|-----------------------|--------|----------------|----------------|---|--|--|
| compd | х | Y | \mathbb{R}^1 | \mathbb{R}^2 | R ³ | VEGFR-2 kinase inhibition ^b | growth inhibition of HUVECs ^c driven by VEGF ^d |
| 3 | S | CH_2 | Н | Н | OCH ₃ | 74^e | 110 |
| 4 | S | CH_2 | Η | Н | CH ₃ | 88^e | ND^{f} |
| 5 | CH_2 | NH | Н | Η | CH ₃ | 280 ^e | ND^{f} |
| 6 | CH_2 | NH | F | Η | CH ₃ | 30 | 20 |
| 7 | CH_2 | NH | F | Н | cyclopropyl | 29 | 14 |
| 8 | CH_2 | CH_2 | F | Н | cyclopropyl | 14 | 5 |
| 9 | CH_2 | NH | F | Η | cyclopentyl | 110 | ND^{f} |
| 10 | CH_2 | NH | F | Н | benzyl | 17 | 120 |
| 11 | CH_2 | NH | F | Н | CH ₂ (2-pyridyl) | 64 | 53 |
| 12 | CH_2 | NH | F | Н | (CH ₂) ₂ -N-morpholine | 85 | ND ^f |
| 13 | CH_2 | NH | F | F | CH ₃ | 71 | 52 |
| 14 | CH_2 | NH | F | F | cyclopropyl | 23 | 25 |

^{*a*} See ref 7b for a description of the assay conditions. ^{*b*} IC₅₀ values are reported as the mean of at least three independent determinations. Variability around the mean value was <50%. ^{*c*} Human umbilical vein endothelial cells. ^{*d*} All compounds evaluated in the HUVEC assay demonstrated an IC₅₀ > 2500 nM in the unstimulated growth of L2987 human lung carcinoma cells. ^{*e*} Single determination. ^{*f*} ND = not determined.

proliferation assays. However, **8** exhibited poor in vitro metabolic stability^{13a} and provided relatively low systemic exposures in mouse 4 h oral PK studies.^{13b} Additional amide analogues, such as the cyclopentyl derivative **9** and **11–12**, which contain terminal polar groups, were 2- to 4-fold less potent than **7** in the VEGFR-2 kinase assay. The benzylamide **10** demonstrated significant enzymatic activity but was nearly one order of magnitude less potent than **7** in the HUVEC proliferation assay.

Additional profiling revealed that the 4-fluorobenzamide derivative **7** possessed low chemical stability under aqueous acidic conditions.^{14a} Thus, a second fluorine atom was introduced on the ring at R^2 in an effort to increase the electron-deficient nature of the benzamide ring. It was postulated that the resulting decrease in basicity of the aniline nitrogen would minimize the potential for acid-catalyzed hydrolysis. Although the corresponding methylamide **13** was slightly less potent in the biochemical assay, cyclopropylamide **14** (BMS-605541) retained excellent in vitro potency and displayed enhanced solution stability at low pH.^{14b} On the basis of its improved chemical stability and impressive in vitro profile, **14** was selected for further characterization in kinase selectivity, PK, and in vivo efficacy studies.

Scheme 1^a



^{*a*} Reagents and conditions: (a) 2-bromo-3-hydroxypropenal, NaOAc, acetone, reflux, 2 h, 66%; (b) (1) thionyl chloride, reflux, 3 h; (2) CH₂Cl₂, cyclopropylamine, Et₃N, -40 °C to room temperature, 80% overall; (c) H₂, 10% Pd/C, EtOAc-EtOH (1:1), 2 h, 97%; (d) Et₃SiH, CH₂Cl₂-TFA (1:1), 1 h, 74%.

| Table 2. | Kinase | Selectivity | Profile | of 14 |
|-----------|----------|-------------|---------|-------|
| I GOIC A. | itiliabe | Delectricy | 1101110 | |

| kinase | enzyme IC50, nM | kinase | enzyme IC ₅₀ , nM |
|----------------|-----------------|-----------------------------------|------------------------------|
| VEGFR-2 | 23 | IKK | >10000 |
| Flk-1 | 40 | SRPK1 | >10000 |
| VEGFR-1 | 400 | IGF-1R | >25000 |
| PDGFR- β | 200 | PKC $\alpha, \delta, \tau, \zeta$ | >40000 |
| c-kit | 370 | Her1 | > 50000 |
| FGFR-1 | >5000 | Her2 | > 50000 |
| GSK-3 | >5000 | Lck | > 50000 |
| MEK | >5000 | Itk | > 50000 |
| AKT | >5000 | Syk | > 50000 |
| p38 | >5000 | JAK-3 | > 50000 |
| Src | >10000 | CaMKII | > 50000 |
| CDK2 | >10000 | MAPKAP K2 | >50000 |

Compound 14 was prepared in a straightforward manner according to the synthetic sequence outlined in Scheme 1. Thus, condensation of thiourea 15^{15} with 2-bromo-3-hydroxypropenal¹⁶ in the presence of sodium acetate in refluxing acetone provided key aldehyde intermediate 16. 5-Amino-*N*-cyclopropyl-2,4-difluorobenzamide (18) was obtained from 2,4-difluoro-5-nitrobenzoic acid 17^{17} using a two-step reaction sequence involving cyclopropylamide bond formation and subsequent reduction of the nitro group. The reductive amination of aldehyde 16 with 18 was accomplished with triethylsilane to afford analogue 14 in 74% yield.

To better understand the enzyme inhibitory properties of 14, the compound was evaluated in an in-house kinase selectivity panel and the K_i was determined for VEGFR-2 kinase. Analogue 14 was confirmed to be an ATP-competitive inhibitor of VEGFR-2, with a K_i of 49 \pm 9 nM. As illustrated in Table 2, 14 was found to be a potent inhibitor of the mouse homologue of VEGFR-2 (Flk-1) and demonstrated moderate activity against VEGFR-1, PDGFR- β , and c-kit. Greater than 200-fold selectivity was observed for all the other receptor and nonreceptor kinase targets found in the panel.

The pharmacokinetic parameters obtained for **14** in mouse, rat, and cynomolgus (cyno) monkey are summarized in Table 3. In each case, **14** demonstrated a moderate steady-state volume of distribution (V_{ss}) and low systemic clearance (Cl) compared to the hepatic blood flow for the respective species. The compound was well absorbed following oral administration of drug from solution formulations at the doses denoted in Table 3. Favorable half-lifes ($t_{1/2}$) and mean residence times (MRT) were observed across species with **14**, albeit these values were slightly higher in cyno. The measured oral bioavailabilities (F_{po}) were 100% for mouse and rat and 52% in monkey.

The encouraging pharmacokinetic profile exhibited by **14** supported additional characterization of the compound in in vivo efficacy studies. Thus, the aminothiazole analogue was evaluated in the L2987 human lung and HCT-116 human colon carcinoma

Table 3. Pharmacokinetic Parameters for 14 in Mouse, Rat, and Cyno

| parameter | mouse ^a | rat ^b | cyno ^b |
|---|--------------------|------------------|-------------------|
| po dose ^c (mg kg ⁻¹) | 90 | 50 | 5 |
| iv dosed (mg kg ⁻¹) | 10 | 10 | 1^e |
| $C_{\max}(\mu M)$, po | 148 | 44.0(6.0) | 8.0(4.0) |
| $T_{\rm max}$ (h), po | 0.5 | 2.0(0) | 0.75(0) |
| AUC (μ M·h), po | 649 ^f | $202^{g}(44)$ | $28.5^{g}(13)$ |
| <i>t</i> _{1/2} (h), po | 1.7 | 2.2(1.2) | 7.1(1.5) |
| MRT (h), po | 3.4 | 3.5(0.4) | 7.9(0.3) |
| Cl (mL min ⁻¹ kg ⁻¹), iv | 11.8 | 13.6(2.1) | 3.9(0.6) |
| $V_{\rm ss}$ (L kg ⁻¹), iv | 1.7 | 1.1(0.1) | 1.6(0.3) |
| $F_{\rm po}$ (%) | 100 | 100 | 52 |

^{*a*} Composition serum concentration-time profiles were constructed for the PK analysis. ^{*b*} Data reported as an average of three animals with associated standard deviations in parentheses. ^{*c*} Vehicle: PEG400/Tween-80/water (40:10:50). ^{*d*} Vehicle: PEG400/Tween-80/water (20:5:75). ^{*e*} Vehicle: 30:70 PEG400/water. ^{*f*} AUC_(0-24h). ^{*g*} AUC_{total}.

 Table 4. In Vivo Antitumor Activity of 14 against Established L2987

 and HCT-116 Xenografts Implanted Subcutaneously in Athymic Mice

| tumor model | dose, mg kg ⁻¹ | schedule | TGI, ^a % |
|----------------------|---------------------------|--------------------|---------------------|
| L2987 ^b | 60 | 1 qd \times 14 | 16 |
| | 90 | | 55 |
| | 180 | | 67 |
| L2987 ^c | 25 | $2qd \times 14$ | 23 |
| | 50 | - | 67 |
| HCT-116 ^c | 12.5 | $2qd \times 14$ | 21 |
| | 25 | - | 51 |
| | 50 | | 64 |

^a Maximum percent tumor growth inhibition over one tumor volume doubling time. ^b Vehicle: PEG400/water (70:30). ^c Vehicle: 100% PEG400.



Figure 2. In vivo efficacy of 14 versus L2987 human tumor xenografts in athymic mice. Circles indicate dosing $(1qd \times 14)$.

xenograft models, and the results are summarized in Table 4.18 In the L2987 study, 14 displayed antitumor activity when administered at doses of 90 and 180 mg/kg once daily for 14 consecutive days (Table 4). Figure 2 illustrates the dosedependent growth inhibition of 14 in the L2987 tumor model. Although considerable tumor growth was noted at the first measurement (day 24), nearly complete tumor stasis was observed at the 180 mg/kg dose level throughout the remainder of the dosing regimen. In an effort to minimize the peak drug concentrations achieved during these studies, a second L2987 tumor xenograft study was conducted at lower doses on a 2qd \times 14 schedule. Thus, twice daily administration of 14 at 50 mg/kg provided an identical TGI as the 180 mg/kg dose level on the once daily schedule (Table 4). Similar TGI values were obtained with 14 in the more sensitive HCT-116 tumor model at doses of 50 and 25 mg/kg on the twice daily dosing regimen. No overt toxicity as measured by weight loss or morbidity was observed at any of the dose levels tested in these experiments.

Structurally related aminothiazole analogues¹² have demonstrated high affinity for the hERG-encoded rapidly activating delayed rectifier potassium channel $(I_{\rm Kr})$.¹⁹ Since potent inhibition of the hERG channel and blockade of the $I_{\rm Kr}$ current have been linked to drug-induced (acquired) QT-interval prolongation and ultimately life threatening ventricular arrhythmias including torsades de pointes, **14** was evaluated in preliminary in vitro cardiovascular safety studies. Compound **14** was found to have an IC₅₀ of 16.6 μ M against the hERG channel expressed in HEK-293 cells.¹⁹ The activity observed in the electrophysiology assay suggests a low to moderate risk for prolongation of the OT interval in vivo.

In conclusion, novel 3-((2-(pyridin-2-ylamino)thiazol-5-ylmethyl)amino)benzamides with potent VEGFR-2 kinase inhibitory activity were identified from the *N*-methoxybenzamidesubstituted pyrrolo[2,1-*f*][1,2,4]triazines using computer-assisted drug design techniques. Optimization of the series SAR led to the identification of **14**, which was orally active in the human lung (L2987) and colon (HCT-116) carcinoma xenograft models at multiple dose levels. On the basis of its favorable in vitro pharmacology, in vivo efficacy, and pharmacokinetic profile, **14** was advanced into preclinical in vivo safety studies.

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Supporting Information Available: Experimental procedures and characterization data for **3–14**, table of combustion analysis and HPLC analysis data for key compounds, and detailed description of pharmacokinetic assays. This material is available free of charge via the Internet at http://pubs.acs.org.

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